

Review

Carbon monoxide – a “new” gaseous modulator of gene expression[★]

Józef Dulak^{1✉} and Alicja Józkowicz^{2,3}

¹*Department of Cell Biochemistry, Faculty of Biotechnology, Jagiellonian University, Kraków, Poland;* ²*Laboratory of Molecular Genetics and Genetic Engineering, Faculty of Biotechnology, Jagiellonian University, Kraków, Poland;* ³*Department of Vascular Surgery, University of Vienna, Austria*

Received: 02 January, 2003; revised 13 February, 2003; accepted: 04 March, 2003

Key words: nitric oxide, oxidative stress, vascular endothelial growth factor, angiogenesis, atherosclerosis

Carbon monoxide (CO) is an odorless, tasteless and colorless gas which is generated by heme oxygenase enzymes (HOs). HOs degrade heme releasing equimolar amounts of CO, iron and biliverdin, which is subsequently reduced to bilirubin. CO shares many properties with nitric oxide (NO), an established cellular messenger. Both CO and NO are involved in neural transmission and modulation of blood vessel function, including their relaxation and inhibition of platelet aggregation. CO, like NO, binds to heme proteins, although CO binds only ferrous (FeII) heme, whereas NO binds both ferrous and ferric (FeIII). CO enhances the activity of guanylate cyclase although it is less potent than NO. In contrast, CO inhibits other heme proteins, such as catalase or cytochrome P450. The effects of CO on gene expression can be thus varied, depending on the cellular microenvironment and the metabolic pathway being influenced. In this review the regulation of gene expression by HO/CO in the cardiovascular system is discussed. Recent data, derived also from our studies, indicate that HO/CO are signifi-

[★]This work was partially supported by grants from the State Committee for Scientific Research (KBN, Poland), No. 3 P04A 049 22 and 6 P04B 013 21.

✉Józef Dulak, Department of Cell Biochemistry, Faculty of Biotechnology, Jagiellonian University, Gronostajowa 7, 30-387 Kraków, Poland; fax: (48 12) 252 6392; e-mail: jdulak@mol.uj.edu.pl

Abbreviations: cGMP, 3',5'-cyclic guanosine monophosphate; HIF, hypoxia inducible factor; ET-1, endothelin-1; HMEC-1, human microvascular endothelial cells; HOs, heme oxygenases; HRE, hypoxia response element; iNOS, inducible nitric oxide synthase; IL-10, interleukin-10; MIF, macrophage inflammatory protein 1 β ; PDGF, platelet-derived growth factor; ROS, reactive oxygen species; SOD, superoxide dismutase; VEGF, vascular endothelial growth factor; TNF- α , tumor necrosis factor α ; VSMC, vascular smooth muscle cells.

cant modulators of inflammatory reactions, influencing the underlying processes such as cell proliferation and production of cytokines and growth factors.

Carbon monoxide has been known since 17th century as a poisonous gas ("the silent killer"). In 1857 Claude Bernard determined that the gas produces asphyxia by reversibly combining with hemoglobin (Piantadosi, 2002). Innumerable deaths have resulted from CO created by incomplete combustion of organic materials. Recently its concentration in the atmosphere, being a result of technological processes and fumes produced by our cars, is monitored and alerts our consciousness when reported to be too high in the smoggy air of our cities. Additionally, CO is a component of cigarette smoke. Due to such a bad reputation not many people, including scientists, are willing to consider its significance as a player of physiological, not only pathological processes in our organism.

Formation of CO in the body was demonstrated in 1952 by Sjöstrand, who reported that decomposition of hemoglobin *in vivo* led to CO production (Sjöstrand, 1952). In 1968 heme oxygenase, the enzymatic source of CO was identified by Tenhunen *et al.* (1968; 1969; 1970), and in the mid 80's two isoforms of the HO enzyme were discovered and cloned (Rotenberg & Maines, 1990; Shibahara *et al.*, 1985; Yoshida *et al.*, 1988). Heme oxygenase-1 (HO-1), an inducible isoform and HO-2, a constitutive form cleave and oxidize the α -methene bridge of the heme molecule yielding equimolar amounts of biliverdin, CO and iron (for a review see: Maines, 1997) (Fig. 1). The catalytic activity of HO requires a concerted action of microsomal NADPH-cytochrome P450 reductase to transfer electrons to the HO-heme complex.

In the last few years evidence has accumulated showing that the HO enzymes and their by-products are important players in the cellular metabolism. This review is intended to discuss some of those data demonstrating the significance of CO in modulation of cellular inflammatory reaction.

HEME OXYGENASES AND THEIR ENZYMATIC ACTIVITY

Currently three isoforms of HO are known: HO-1, HO-2 and HO-3. HO-1 (termed also hsp32) is a stress inducible enzyme which can be expressed most probably in every cell facing contact with noxious stimuli (for a review see: Maines, 1997). Hence, HO-1 induction can be regarded as a general response to oxidant stress (Applegate *et al.*, 1991). HO-1 can represent a secondary protective system, while the primary defense mechanism against oxidative stress is glutathione (Meister, 1994). Interestingly, glutathione depletion results in a strong induction of HO-1 (Applegate *et al.*, 1991; Lautier *et al.*, 1992; Ewing & Maines, 1993) supporting the importance of HO-1 in cellular protective mechanisms.

HO-2 is a constitutive gene, expressed in neurons, endothelium and many other cell types. The only known inducers of HO-2 activity are adrenal glucocorticoids (Weber *et al.*, 1994) but it is also possible that activation of protein kinase C can result in an increased degradation of heme by the HO-2 isoform (Baranano & Snyder, 2001). HO-2 is involved in the regulation of neural system functioning, modulating the neural transmission in central nervous system, digestive system and in male copulatory organ (for a review see: Maines, 1997; Baranano & Snyder, 2001; Snyder & Baranano, 2001).

HO-3 is a newly identified isoform (McCoubrey *et al.*, 1997) and has been found only in rats (Scapagnini *et al.*, 2002). It is constitutively expressed in the liver, spleen, brain and kidney, but its ability to degrade heme is much limited in comparison with HO-1 and HO-2 (McCoubrey *et al.*, 1997). The HO-3 gene does not contain introns which suggests that HO-3 could have arisen by retrotransposition of the HO-2 gene (Scapagnini *et al.*, 2002). HO-3 is believed to function as a heme sensing

or heme binding protein rather than a heme-degrading enzyme (McCoubrey *et al.*, 1997), but its biological role requires further elucidation.

The activity of HO-1 generates colour effects. When large amounts of heme are released from destroyed erythrocytes due to a physical insult on blood vessels in the skin and muscles, HO-1, induced by heme, starts its degradation. The black heme is transformed to green biliverdin, and when the bruise is disappearing, the yellowish bilirubin appears.

Basing on the biochemical activities of HO, its products have for a long time been recognized only as the waste of heme cleavage. Additionally, when heme degradation occurs under pathological conditions, significant health disturbances may arise. Hence, it has been customary to regard HO as an enzyme, the activity of which results in the clinical problem of hyperbilirubinemia leading to jaundice. However, ample evidence has recently accumulated indicating that HO activity and all its by-products play important physiological roles (for reviews see: Choi & Otterbein, 2002; Foresti & Motterlini, 1999; Hill-Kapturczak *et al.*, 2002; Maines, 1997; Otterbein & Choi, 2000) (Fig. 1).

First, HO activity removes the prooxidant heme (Balla *et al.*, 1991; Jeney *et al.*, 2002; Quan *et al.*, 2002; Vercellotti *et al.*, 1994). Second, both biliverdin and bilirubin have antioxidant properties, efficiently scavenging reactive oxygen species and inhibiting lipid peroxidation (Baranano *et al.*, 2002; Stocker *et al.*, 1987a; 1987b). Third, iron released from heme enhances the synthesis of ferritin, which additionally has antioxidant capabilities (Balla *et al.*, 1992). Fourth, HOs, or at least HO-1, prevent free iron accumulation in the cells not only indirectly by stimulating ferritin production, but also directly by extruding iron outside the cell (Ferris *et al.*, 1999). In this activity HO-1 cooperates with a recently identified Fe-ATP pump (Baranano *et al.*, 2000). An indication for a role of HO-1 in iron extrusion is the accumulation of iron in the cells of *HO-1* knockout mice (Poss & Tonegawa, 1997b; 1997c) or in the liver and kidney of a boy who lacked a functional *HO-1* gene (Yachie *et al.*, 1999). Accordingly, gene transfer of *HO-1* to cells derived from *HO-1* knockout animals restored the cells' capability to control the cellular iron level (Ferris *et al.*, 1999).

Finally, CO, the "notoriously infamous" by-product of HO activity enters recently the

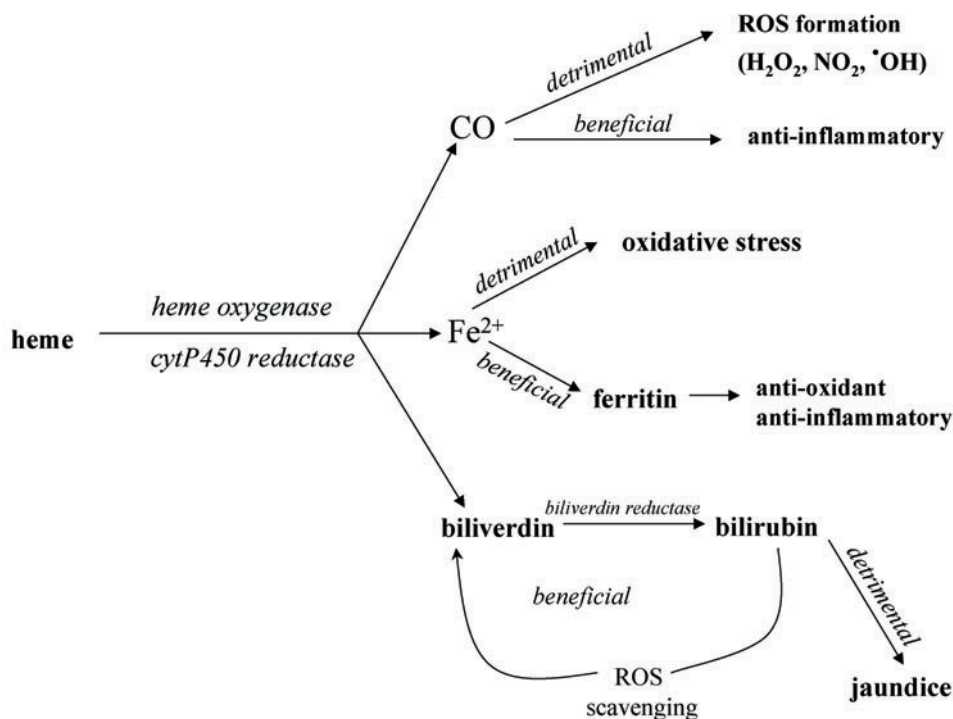


Figure 1. Mechanisms of heme oxygenase activity

scene as an important modulator of many physiological processes. Particularly, it plays a role in neural transmission, is necessary for ejaculation, involved in the homeostatic control of cardiovascular function and appears to modulate gene expression in many cell types (for reviews see: Foresti & Motterlini, 1999; Snyder & Baranano, 2001). However, due to space limitations we will discuss here only the recently investigated activities of CO in the pathways governing the synthesis of inflammatory cytokines and growth factors.

CO PRODUCTION AND MECHANISMS OF CO EFFECTS ON CELLULAR FUNCTION

HOs are the main producers of CO in the human body. Much smaller amounts of CO can also derive from other sources, like lipid peroxidation (Fig. 2) (for a review see: Piantadosi, 2002).

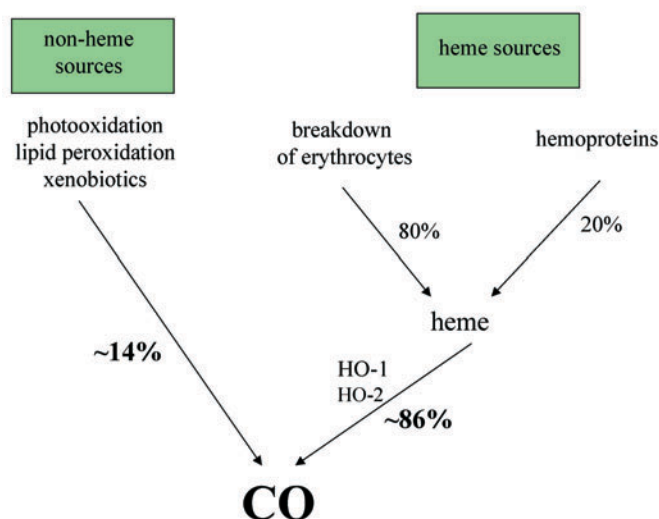


Figure 2. The sources of carbon monoxide (based on data from Vreman *et al.*, 2002)

tadosi, 2002). CO is formed at the rate of 16.4 $\mu\text{mol/h}$ in the human body and the daily production of CO is substantial, reaching more than 12 ml (500 μmole). However, average physiological concentrations of CO in tissues are rather in the low nanomolar range (for a review see: Piantadosi, 2002). On the other hand, increased production of CO has been demonstrated in chronic inflammatory lung diseases, such as obstructive pulmonary dis-

ease, cystic fibrosis, and asthma as well as in infectious pulmonary diseases (Kharitonov & Barnes, 2002).

The action of CO depends primarily on its ability to bind heme proteins and to inhibit or alter their biochemical functions (for a review see: Piantadosi, 2002). By interacting with heme proteins CO differentially influences electron-transport reactions producing either prooxidant or antioxidant effects. The action of CO is dependent on its concentration, the concentration of O_2 as well as on the availability of reduced transition metals, such as iron or copper (cited after: Piantadosi, 2002).

Some activities of CO resembles those of NO (for reviews see: Baranano & Snyder, 2001; Snyder *et al.*, 1998). CO, like NO, avidly binds heme proteins with iron in the reduced ferrous state [Fe(II)], but unlike NO, does not bind ferric iron [Fe(III)]. Like NO, CO activates soluble guanylate cyclase (Brune *et al.*, 1990; Furchgott & Jothianandan, 1991;

Karlsson *et al.*, 1985; Utz & Ullrich, 1991), leading to a several-fold increase in the production of cGMP, although its potency in stimulation of this effect is about 30–100 times lower than that of NO (Kharitonov *et al.*, 1995). Therefore, due to such a weak influence of CO on guanylate cyclase, the physiological significance of this activation is in doubt. It is, however, suggested that the effect of CO on guanylate cyclase can be potentiated

by some as yet unknown co-stimulators. The rationale for such a hypothesis was a discovery of YC-1, a benzyloid derivative, which augments CO-mediated induction of cGMP production to a level attained by NO (Friebe & Koesling, 1998; Friebe *et al.*, 1996).

Additionally, it is possible that CO may amplify the NO-mediated activation of guanylate cyclase (Ingi *et al.*, 1996; Cao *et al.*, 2000). CO may also exert its activity through a direct influence on NO. Indeed, low concentrations of CO stimulate NO release and augment the production of the strong oxidant peroxynitrite in vascular cells (Thom *et al.*, 1999; 2000).

Other heme proteins, such as myoglobin, cytochrome *c* oxidase, cytochrome P450, catalase and tryptophan dioxygenase can be also influenced by CO (for a review see: Piantadosi, 2002). In contrast to guanylate cyclase, however, CO inhibits their catalytic activities. As a result, a significant cellular oxidative stress can be produced by CO in vascular endothelium (Thom *et al.*, 1999; 2000) and in other cell types *in vivo* (Piantadosi, 2002). This occurs already after low-level CO exposure (100 p.p.m.) and can generate a significant lipid peroxidation. Such an effect can be blocked by superoxide dismutase and iron chelators (Piantadosi, 2002).

CO can influence gene expression in several ways. First, an increase in CO concentration *in vivo* will result in carboxyhemoglobin formation and decreased oxygenation and hypoxia. Hypoxia is a physiological regulator of important biological processes, including erythropoiesis, angiogenesis, glycolysis and tissue remodeling (for a review see: Kourembanas, 2002). It may originate from a decrease in O₂ concentration, but can also occur when blood hemoglobin is blocked by CO.

Second, local effects of CO may derive from its interaction with NO (for a review see: Hartsfield, 2002). CO can cause a release of NO from its heme-bound intracellular pool, which may result not only in activation of guanylate cyclase, but also in nitrosylation of protein thiol groups (Thorup *et al.*, 1999;

Foresti *et al.*, 1999; Marshall *et al.*, 2000). Through those ways NO can influence gene expression (for reviews see: Dulak & Józkwicz, 2003; Marshall *et al.*, 2000). Such effects can occur either in cells constitutively generating NO, like neurons and endothelial cells, or in cells producing significant amounts of NO by the action of inducible nitric oxide synthase (iNOS). A combination of high concentrations of NO and CO can influence mitochondrial cytochromes, causing their inhibition and formation of reactive oxygen species (ROS) leading to lipid peroxidation (Agarwal *et al.*, 1995; Koehler & Traystman, 2002). ROS induced by CO can influence the activity of several transcription factors and kinases, as has been demonstrated so far for NF- κ B and p38 kinase (Brouard *et al.*, 2002) (see below).

Third, the cellular effect of CO can be independent of hypoxia and interaction with NO. As mentioned, CO can induce the generation of hydrogen peroxide, maybe through induction of the expression of manganese superoxide dismutase (MnSOD) (Frankel *et al.*, 2000) or by inhibition of catalase activity (Zhang & Piantadosi, 1992; Piantadosi, 2002) (Fig. 3). H₂O₂ is not only a toxic oxidant, but it is also an important cellular messenger, regulating the expression of numerous genes.

Besides the similarities striking differences between CO and NO exist. NO is a free radical and it is the most reactive of physiological gases (Piantadosi, 2002). The reaction of NO with ROS can result in formation of numerous potent intermediates. It is likely that NO toxicity, which is higher than that of CO and occurs above 100 p.p.m., derives from its reaction with superoxide to form peroxynitrite (Piantadosi, 2002). Accordingly, the potential therapeutic doses of inhaled NO are probably lower (65 p.p.m.) than of carbon monoxide (500–1000 p.p.m.) (Thiemermann, 2001).

In vivo CO is almost immediately toxic at the concentration of 0.4% (4000 p.p.m.) or more, but the concentration of 0.01% (100 p.p.m.) is tolerable and allowable for an exposure of several hours (Otterbein & Choi, 2000). CO at low

concentrations (10–500 p.p.m.) is well tolerated by cells, and the rodents can be exposed to 500 p.p.m. continuously for up to 2 yr without deleterious effects (Stupfel & Bouley, 1970; Otterbein & Choi, 2000).

The cellular effects of CO was studied basing on the activity of HO-1 and the use of HO inhibitors, but data on cellular CO concentrations are usually lacking. In contrast, exogenous CO was applied at very different amounts, sometimes very high. Such varied modes of treatment may result in discrepant results as will be discussed further.

REGULATION OF GENE EXPRESSION IN HYPOXIA BY CO

In vascular smooth muscle cells (VSMC) cultured under hypoxic conditions CO is produced as a result of induction of HO-1, and an increase in cGMP content is observed (Morita *et al.*, 1995). The response is transient, with cGMP peaking at 15 h and returning to baseline by 48 h. It has been reported that very high exogenous CO concentrations (5% or more, even up to 80%) inhibited hypoxic induction of erythropoietin (Huang *et al.*, 1999), vascular endothelial growth factor (VEGF) (Goldberg & Schneider, 1994), endothelin-1 (ET-1) and platelet-derived growth factor (PDGF) genes (Morita & Kourembanas, 1995). Endogenous CO, derived from HO activity in VSMC growing in co-culture with endothelial cells, inhibited the hypoxic induction of PDGF-B and ET-1 in endothelium (Morita & Kourembanas, 1995). In a feedback response, the decreased production of those mitogens by endothelial cells resulted in slowing down the proliferation of the co-cultured VSMC (Morita *et al.*, 1997).

The underlying mechanism is not known. It was suggested that it can be due to the inhibition of cytochrome P450 (Wang, 1998). Indeed, cytochrome P450-linked monooxygenase is responsible for the generation of vasoconstricting substances, such as certain

arachidonic acid metabolites or ET-1. CO is an inhibitor of cytochrome P450 and the level of cytochrome P450 is controlled by the availability of cellular heme, which can be degraded to CO and biliverdin (Wang, 1998). A decreased formation of vasoconstrictors in response to CO would lead to vascular relaxation.

The complex effect of CO on gene expression in hypoxia requires elucidation. It has been hypothesized that CO inhibits the hypoxic induction of genes encoding vasoconstrictors in smooth muscle cells in the early hypoxic phase (Kourembanas, 2002). During chronic hypoxia, however, low CO may tilt the balance toward increased production of growth factors and vasoconstrictors that promote vessel-wall remodeling (Kourembanas, 2002).

The main molecular sensor of the level of O₂ in the cell is the transcription factor HIF-1 (for a review see: Semenza, 2002). In the functional state it is a heterodimer consisting of two subunits, HIF-1 α and HIF-1 β . This heterodimer binds to a recognition site (HRE – hypoxia response element) present in the promoter of many genes regulated by hypoxia, such as erythropoietin, VEGF, HO-1 or inducible nitric oxide synthase (iNOS) (for a review see: Semenza, 2002).

HIF-1 subunits are constitutively produced in the majority of cells, but dimer formation is prevented by oxygen-dependent degradation of HIF-1 α . Degradation occurs after hydroxylation of specific proline residues (P402 and P564) in the HIF-1 α protein (for a review see: Maxwell & Ratcliffe, 2002). At normal oxygen tension the hydroxylation is performed by a prolyl hydroxylase, which requires iron, O₂ and 2-oxoglutarate as cofactors.

The data on the effect of CO on HIF-1 production and activity are scarce and inconclusive. It has been suggested that the cellular hypoxia sensor is a heme-containing protein (Goldberg & Schneider, 1994; Huang *et al.*, 1999) because cobalt chloride or iron chelators can mimic the effects of hypoxia. It has been hypothesized that in the presence of

O₂ that putative protein can bind O₂ at a heme site attaining a “relaxed” configuration, whereas the absence of O₂ confers a “tense” conformation (cited after Kourambanas, 2002). It was claimed that CO, a molecule known to interact with heme groups, can inhibit the hypoxic induction of genes by behaving similarly to O₂ and shifting the heme-protein to the relaxed configuration (Kourambanas, 2002). However, the recent discovery of the prolyl hydroxylase which requires iron but is not a heme protein sheds some doubts on this attractive hypothesis.

Recently, a first eukaryotic transcription factor selectively affected by CO was discovered. Dioum *et al.* (2002) have demonstrated that binding of NPAS2, a member of the same family of proteins to which HIF-1 α belongs, is inhibited by CO, but is not influenced by NO nor O₂. NPAS2 is a homologue of CLOCK, a transcription factor involved in modulation of circadian activity in the suprachiasmatic nucleus (Rutter *et al.*, 2002). NPAS2 is also present in cells outside the central nervous system (Dioum *et al.*, 2002; Rutter *et al.*, 2002), and, interestingly, binds to a very similar DNA sequence (CACGTG) as HIF-1 (TACGTG). The relationship and potential reciprocal influence of NPAS2 and HIF-1 on their activity is not known and opens a new fascinating area for further investigations.

The binding of the HIF-1 protein to HRE, as determined by gel shift assay, was attenuated in cells treated with exogenous CO in hypoxia (Liu *et al.*, 1998). However, the amount of HIF-1 α protein seemed not to be influenced at 5% CO concentration (Liu *et al.*, 1998), while it was decreased at high, 80% exposure (Huang *et al.*, 1999). How this relates to the physiological situation is not known as the concentrations of CO used in those experiments were extremely high.

Interestingly, and in contrast to the studies discussed above, our recent data indicate that CO can be a positive regulator of VEGF synthesis. We observed a significant induction of

VEGF in vascular smooth muscle cells cultured in the presence of 1% CO (10000 p.p.m.) in otherwise normoxic conditions (Dulak *et al.*, 2002; Dulak & Józkwicz, 2003). It appears that CO can modulate VEGF synthesis also in other cell types. Accordingly, we observed an increase in VEGF synthesis in microvascular endothelial cells (HMEC-1) treated with ruthenium carbonyl compound, a representative of a new class of substances named carbon monoxide releasing molecules (CO-RM) (Jozkwicz *et al.*, in press). Those chemicals can release CO, and thus are equivalent to the widely used NO donors (Motterlini *et al.*, 2002). Additionally, in HMEC-1 the expression of VEGF was more potently increased by 15d-PGJ₂, a strong activator of HO-1, which induces the generation of higher amount of CO than that attained after CO-RM treatment (Jozkwicz *et al.*, in press).

THE EFFECT OF CO ON INFLAMMATORY REACTION

Great attention has been recently paid to the presumed anti-inflammatory functions of CO. It has been demonstrated that CO at a physiological concentration (100–500 p.p.m.) inhibits the production of pro-inflammatory cytokines (Otterbein *et al.*, 2000). In macrophages treated with LPS the synthesis of TNF α , MIF and IL-1 is a marker of the inflammatory processes. When such cells were kept in the presence of CO, the production of those pro-inflammatory molecules decreased. Interestingly, CO upregulated the synthesis of the anti-inflammatory cytokine IL-10 (Otterbein *et al.*, 2000). Looking further for the mechanisms governing this potentially preventive CO activity Otterbein *et al.* (2000) found that CO action is independent of cGMP, but rather p38 kinase is necessary (Fig. 3). Accordingly, CO failed to inhibit cytokine production in cells derived from animals with targeted mutation in the

MKK3 gene encoding a kinase activating p38 (Otterbein *et al.*, 2000).

Interestingly, a positive loop may operate in the anti-inflammatory functions of HO-1. As mentioned, CO induces IL-10 synthesis in monocytes, indicating for a role of HO-1. Recently Lee & Chau (2002) have demonstrated that IL-10 induces HO-1 expression, utilizing CO as a mediator of anti-inflammatory activities.

The anti-inflammatory activity of CO may underlie the protective effect of HO in such processes like graft rejection (Ke *et al.*, 2001; 2002; Sato *et al.*, 2001) or development of atherosclerosis (Ishikawa & Maruyama, 2001; Ishikawa *et al.*, 2001). It has been elegantly demonstrated that HO activity is necessary for inhibition of xenograft or allograft rejection. When hearts are transplanted from mice to rats, the acute rejection can be prevented by ablation of the complement system and suppression immune response in the recipients by cobra venom factor treatment and cyclosporine delivery (Soares *et al.*, 1998). However, when HO activity was additionally blocked by HO inhibitors the grafts were rejected (Soares *et al.*, 1998).

Through the influence on inflammatory reactions HO-1 overexpression may also prevent the development of atherosclerosis (Shi

et al., 2000). The beneficial effect of HO-1 can be exerted by CO-dependent inhibition of vessel constriction (Suematsu *et al.*, 1995), while bilirubin can attenuate the adhesiveness of leukocytes to the vessel wall (Duckers *et al.*, 2001; Hayashi *et al.*, 1999; Kozma *et al.*, 1997; 1999; Zhang *et al.*, 2001). Additionally, HO-1 activity may modulate the inflammatory processes by augmenting iron extrusion from the cells of the blood vessel wall (Juan *et al.*, 2001).

Disruption of the HO-1 gene provides very interesting data supporting the anti-inflammatory role of HO products. The life of HO-1 knockout mice is strongly affected by progressive chronic inflammation characterized by hepatosplenomegaly, lymphadenopathy and leukocytosis. Animals are debilitated and die at a young age from massive iron overload in the liver and kidneys (Poss & Tonegawa, 1997a; 1997b). In response to chronic hypoxia, they exhibit enhanced lipid peroxidation, accentuated oxidative damages, and increased right ventricular infarcts with organized mural thrombi. Exposure to endotoxin results in strong hepatocellular necrosis and high mortality from endotoxic shock. The cells derived from HO-1 targeted mice are highly susceptible to heme- and hydrogen peroxide-mediated toxicity. HO-1 knockout mice are

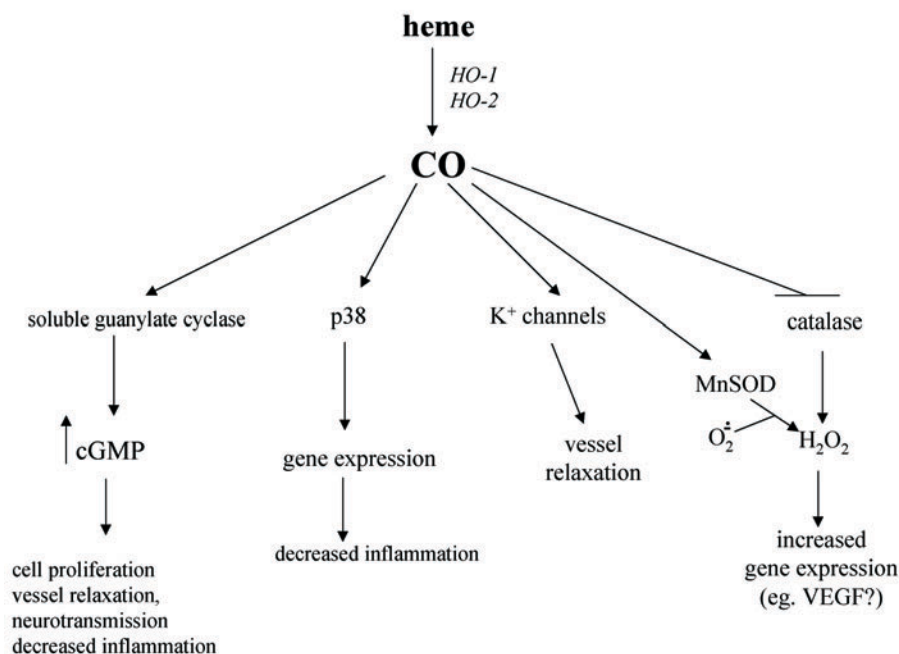


Figure 3. The effect of CO on gene expression and physiological functions in the cardiovascular system.

In this figure only the potential beneficial effects of CO are mentioned. However, it remains to be established whether the detrimental effects, which can occur when HO-1 activity is very high (Suttner & Dennery, 1999), are solely due to the increased release of free iron or are also dependent on CO.

also more sensitive to pulmonary ischemia compared to wild type counterparts (Fujita *et al.*, 2001). This could be prevented by inhalation of CO (0.1%), which inhibited the hypoxic induction of plasminogen activator inhibitor (PAI-1), resulting in higher activity of tissue plasminogen activator (tPA) and decreased fibrin deposition and lung inflammation (Fujita *et al.*, 2001).

Interestingly, the first case of human HO-1 deficiency was described after initial studies on HO-1 targeted mice. The human case of HO-1 deficiency exhibited similar features, including iron deposition in renal and hepatic tissues (Kawashima *et al.*, 2002; Ohta *et al.*, 2000; Yachie *et al.*, 1999). Lack of HO-1 activity resulted in extreme vulnerability of vessels to common stressful stimuli, including infections and environmental toxic substances. A cascade of inflammatory reactions and sustained oxidative stress led to severe and persistent vascular endothelial damage and detachment. Cell lines derived from this patient were strongly sensitive to hemin-induced injury (Jeney *et al.*, 2002). Importantly, HO-2, which was expressed normally, did not provide efficient defense. These clinical symptoms demonstrate the critical importance of HO-1 rather than HO-2 in iron metabolism and in protection of vessels against oxidative stress.

THE INHIBITORY EFFECT OF CO ON CELL PROLIFERATION AND APOPTOSIS

Growth factors, such as PDGF-BB (Durante *et al.*, 1999) or transforming growth factor β (TGF- β) (Kutty *et al.*, 1994; Hill-Kapturczak *et al.*, 2000) which induce VSMC proliferation, enhance also HO-1 expression. It has been hypothesized that induction of HO-1 expression represents a counterbalanced autocrine mechanisms which limits the SMC proliferation (Kourembanas, 2002). Indeed, gene transfer of HO-1 (Duckers *et al.*, 2001; Liu *et al.*, 2002;

Peyton *et al.*, 2002; Zhang *et al.*, 2002) or delivery of CO (Morita *et al.*, 1997) blocked SMC proliferation. Accordingly, inhibition of HO-1 activity potentiates SMC growth (Peyton *et al.*, 2002; Togane *et al.*, 2000). The same effect was obtained by treatment of growing SMC with hemoglobin, a CO scavenger (Peyton *et al.*, 2002).

The effect of CO on the proliferation of VSMC can be mediated by influence on transcription factors. Among them E2F is a family of cell-cycle specific transcription factors, which regulate the expression of many genes involved in cell proliferation, and govern the transition of cells from the G₁ to S phase. It has been demonstrated that E2F is affected by CO in a cGMP-dependent manner (Durante, 2002). The involvement of cGMP in the inhibition of VSMC proliferation was also corroborated by experiments with YC-1, which augments CO-dependent activation of guanylate cyclase (Friebe & Koesling, 1998). Accordingly, YC-1 attenuated the growth of smooth muscle cells (Durante, 2002). Moreover, inhibition of guanylate cyclase with methylene blue or ODQ ([1,2,4]-oxadiazolo[4,3- α] quinoxaline-1-one) prevented CO effect on SMC cell cycle progression (Morita *et al.*, 1997).

The inhibition of the growth of VSMC can have important consequences for prevention of the vessel narrowing after balloon angioplasty, a therapeutic treatment applied to patients with atherosclerosis developing in their coronary peripheral vessels. It is presumed that quick regeneration of endothelial cells after angioplasty can inhibit restenosis due to the restoration of endogenous modulatory mechanisms governing vessel functions (for review see: Dulak & Józkwicz, 2002). Interestingly, it has been recently demonstrated that CO protects endothelial cells from apoptosis induced by various stimuli (Brouard *et al.*, 2000). This effect requires the activation of the NF- κ B transcription factor and is dependent on p38 kinase activity (Brouard *et al.*, 2002) indicating for a common mechanism of

the protective action of CO in endotoxin- or cytokine-induced inflammation and in regenerative processes after mechanical injury.

Of particular interest are observations demonstrating that CO (or HO-1 transfection) enhanced proliferation of endothelial cells (Li Volti *et al.*, 2002; Malaguarnera *et al.*, 2002; Deramaudt *et al.*, 1999). Thus, CO may behave similarly like NO, which prevents the apoptosis of endothelial cells and stimulates their proliferation, but which inhibits the growth of SMC (Kibbe *et al.*, 2000; Tzeng *et al.*, 1997). The mechanisms of such effects of those gases on endothelial cells are not known. It can be speculated that both CO and NO enhance the expression of VEGF, a mitogen and anti-apoptotic factor for endothelial cells (Dulak, 2001; Dulak *et al.*, 2002; Józkowicz *et al.*, in press). However, as endothelial cells do not always produce detectable quantities of VEGF, other mechanisms may lay behind the effect of CO in endothelial cells.

Again, contradictory data have been also obtained concerning the effect of CO on viability of endothelial and vascular smooth muscle cells. Thom *et al.* (2000) showed that maintaining of bovine pulmonary artery endothelial cells in the presence of 100 p.p.m. CO for more than 1 h caused cell death, which could be prevented by a caspase-1 inhibitor. The effect of CO was mediated by NO, as it was blocked by an NO synthase inhibitor, S-isopropylisothiourea, and the peroxynitrite scavenger selenomethionine. Interestingly, prior exposure of endothelial cells to a lower concentration of CO, 10 p.p.m., conferred resistance against the lethal effects of 100 p.p.m. CO, suggesting that the anti- and pro-apoptotic effect of CO is concentration dependent (Thom *et al.*, 2000).

CONCLUSIONS

There is no doubt that CO exerts significant effects on many pathways of the cellular metabolism. In cells of the cardiovascular system

CO inhibits inflammatory response, influencing synthesis of cytokines, cell proliferation and preventing cell apoptosis (Fig. 3). Those effects are mediated through both cGMP-dependent and cGMP-independent ways. The physiological activity of CO may result in inhibition of inflammatory reactions in hyperoxia, ischemia/reperfusion injury, atherosclerosis and graft rejection.

It cannot be longer taken as a dogma that CO is only a deadly substance with no physiological functions. Rather, CO can be regarded as a signaling molecule involved in, and maybe even critical for many aspects of cellular metabolism. Apparently, like in the case of the majority of substances, its potentially harmful or beneficial effects are dependent on the concentration. Regarding the physiological role, low concentration of CO, equivalent to the amount released by local activity of heme oxygenase, can influence underlying processes, inhibiting the inflammatory reactions. However, the range of HO activity is much broader, as it concomitantly releases biliverdin and iron. The final outcome of HO activity is thus probably different than the effect expected from the action of separately delivered CO. The beneficial, and possibly therapeutic window of HO-1 appears to be quite narrow, with protective effects exerted at moderately increased activity, and harmful influences prevailing when iron release from degraded heme is very high (Dennery *et al.*, 2003; Suttner & Dennery, 1999). Therefore, the potential beneficial and even therapeutic effects ascribed to increased HO-1 expression or CO supplementation have to be carefully reconsidered regarding the hypothetical risk of aggravation of the oxidative stress due to the increased CO and free iron release.

Although the complexity of the mechanisms underlying CO actions on gene expression is not well known, the results obtained in the last few years have demonstrated its importance in modulation of inflammatory reaction and cellular growth. They shed light on many unknown aspects of CO functions and suggest

new avenues for further investigations and presumably therapeutic applications.

We are grateful to Professor Aleksander Koj for his comments.

REFERENCES

- Agarwal A, Balla J, Alam J, Croatt AJ, Nath KA. (1995) Induction of heme oxygenase in toxic renal injury: a protective role in cisplatin nephrotoxicity in the rat. *Kidney Int.*; **48**: 1298–307.
- Applegate LA, Luscher P, Tyrrell RM. (1991) Induction of heme oxygenase: a general response to oxidant stress in cultured mammalian cells. *Cancer Res.*; **51**: 974–8.
- Balla G, Jacob HS, Balla J, Rosenberg M, Nath K, Apple F, Eaton JW, Vercellotti GM. (1992) Ferritin: a cytoprotective antioxidant strategem of endothelium. *J Biol Chem.*; **267**: 18148–53.
- Balla G, Vercellotti GM, Muller-Eberhard U, Eaton J, Jacob HS. (1991) Exposure of endothelial cells to free heme potentiates damage mediated by granulocytes and toxic oxygen species. *Lab Invest.*; **64**: 648–55.
- Baranano DE, Snyder SH. (2001) Neural roles for heme oxygenase: contrasts to nitric oxide synthase. *Proc Natl Acad Sci U S A.*; **98**: 10996–1002.
- Baranano DE., Rao M., Ferris CD, Snyder SH. (2002) Biliverdin reductase: a major physiologic cytoprotectant. *Proc Natl Acad Sci U S A.*; **99**: 16093–8.
- Baranano DE, Wolosker H, Bae BI, Barrow RK, Snyder SH, Ferris CD. (2000) A mammalian iron ATPase induced by iron. *J Biol Chem.*; **275**: 15166–73.
- Brouard S, Berberat PO, Tobiasch E, Seldon MP, Bach FH, Soares MP. (2002) Heme oxygenase-1-derived carbon monoxide requires the activation of transcription factor NF-kappa B to protect endothelial cells from tumor necrosis factor-alpha-mediated apoptosis. *J Biol Chem.*; **277**: 17950–61.
- Brouard S, Otterbein LE, Anrather J, Tobiasch E, Bach FH, Choi AM, Soares MP. (2000) Carbon monoxide generated by heme oxygenase 1 suppresses endothelial cell apoptosis. *J Exp Med.*; **192**: 1015–26.
- Brune B, Schmidt KU, Ullrich V. (1990) Activation of soluble guanylate cyclase by carbon monoxide and inhibition by superoxide anion. *Eur J Biochem.*; **192**: 683–8.
- Cao L, Blute TA, Eldred WD. (2000) Localization of heme oxygenase-2 and modulation of cGMP levels by carbon monoxide and/or nitric oxide in the retina. *Vis Neurosci.*; **17**: 319–29.
- Choi AM, Otterbein LE. (2002) Emerging role of carbon monoxide in physiologic and pathophysiologic states. *Antioxid Redox Signal.*; **4**: 227–8.
- Dennery PA, Visner G, Weng YH, Nguyen X, Lu F, Zander D, Yang G. (2003) Resistance to hyperoxia with heme oxygenase-1 disruption: role of iron. *Free Radical Biol Med.*; **34**: 124–33.
- Deramaudt BM, Braunstein S, Remy P, Abraham NG. (1998) Gene transfer of human heme oxygenase into coronary endothelial cells potentially promotes angiogenesis. *J Cell Biochem.*; **68**: 121–7.
- Deramaudt BM, Remy P, Abraham NG. (1999) Upregulation of human heme oxygenase gene expression by Ets-family proteins. *J Cell Biochem.*; **72**: 311–21.
- Dioum EM, Rutter J, Tuckerman JR, Gonzalez G, Gilles-Gonzalez MA, McKnight SL. (2002) NPAS2: a gas-responsive transcription factor. *Science.*; **298**: 2385–7.
- Duckers HJ, Boehm M, True AL, Yet SF, San H, Park JL, Clinton Webb R, Lee ME, Nabel GJ, Nabel EG. (2001) Heme oxygenase-1 protects against vascular constriction and proliferation. *Nat Med.*; **7**: 693–8.
- Dulak J. (2001) New mechanisms of regulation of vascular endothelial growth factor synthesis. The role of nitric oxide and carbon mon-

- oxide. *Habilitation thesis*. Institute of Molecular Biology, Jagiellonian University, Krakow.
- Dulak J, Józkowicz A. (2002) Angiogenic gene therapy with vascular endothelial growth factor: hope or hype? *Eur Surg.*; **34**: 101–4.
- Dulak J, Józkowicz A. (2003) Regulation of vascular endothelial growth factor synthesis by nitric oxide: facts and controversies. *Antioxid Redox Signal.*; **5**: 123–32.
- Dulak J, Jozkowicz A, Foresti R, Kasza A, Frick M, Huk I, Green CJ, Pachinger O, Weidinger F, Motterlini R. (2002) Heme oxygenase activity modulates vascular endothelial growth factor synthesis in vascular smooth muscle cells. *Antioxid Redox Signal.*; **4**: 229–40.
- Durante W. (2002) Carbon monoxide and vascular smooth muscle cell growth. In *Carbon monoxide and cardiovascular functions*. Wang R, ed, pp 45–65. CRC Press, Boca Raton, FL.
- Durante W, Peyton KJ, Schafer AI. (1999) Platelet-derived growth factor stimulates heme oxygenase-1 gene expression and carbon monoxide production in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol.*; **19**: 2666–72.
- Ewing JF, Maines MD. (1993) Glutathione depletion induces heme oxygenase-1 (HSP32) mRNA and protein in rat brain. *J Neurochem.*; **60**: 1512–9.
- Ferris CD, Jaffrey SR, Sawa A, Takahashi M, Brady SD, Barrow RK, Tysoe SA, Wolosker H, Baranano DE, Dore S, Poss KD, Snyder SH. (1999) Haem oxygenase-1 prevents cell death by regulating cellular iron. *Nat Cell Biol.*; **1**: 152–7.
- Foresti R, Motterlini R. (1999) The heme oxygenase pathway and its interaction with nitric oxide in the control of cellular homeostasis. *Free Radical Res.*; **31**: 459–75.
- Frankel D, Mehindate K, Schipper HM. (2000) Role of heme oxygenase-1 in the regulation of manganese superoxide dismutase gene expression in oxidatively-challenged astroglia. *J Cell Physiol.*; **185**: 80–6.
- Friebe A, Koesling D. (1998) Mechanism of YC-1-induced activation of soluble guanylyl cyclase. *Mol Pharmacol.*; **53**: 123–7.
- Friebe A, Schultz G, Koesling D. (1996) Sensitizing soluble guanylyl cyclase to become a highly CO-sensitive enzyme. *EMBO J.*; **15**: 6863–8.
- Fujita T, Toda K, Karimova A, Yan SF, Naka Y, Yet SF, Pinsky DJ. (2001) Paradoxical rescue from ischemic lung injury by inhaled carbon monoxide driven by derepression of fibrinolysis. *Nat Med.*; **7**: 598–604.
- Furchgott RF, Jothianandan D. (1991) Endothelium-dependent and -independent vasodilation involving cyclic GMP: relaxation induced by nitric oxide, carbon monoxide and light. *Blood Vessels.*; **28**: 52–61.
- Goldberg MA, Schneider TJ. (1994) Similarities between the oxygen-sensing mechanisms regulating the expression of vascular endothelial growth factor and erythropoietin. *J Biol Chem.*; **269**: 4355–9.
- Hancock WW, Buelow R, Sayegh MH, Turka LA. (1998) Antibody-induced transplant arteriosclerosis is prevented by graft expression of anti-oxidant and anti-apoptotic genes. *Nat Med.*; **4**: 1392–6.
- Hartsfield CL. (2002) Cross talk between carbon monoxide and nitric oxide. *Antioxid Redox Signal.*; **4**: 301–7.
- Hayashi S, Takamiya R, Yamaguchi T, Matsumoto K, Tojo SJ, Tamatani T, Kitajima M, Makino N, Ishimura Y, Suematsu M. (1999) Induction of heme oxygenase-1 suppresses venular leukocyte adhesion elicited by oxidative stress: role of bilirubin generated by the enzyme. *Circ Res.*; **85**: 663–71.
- Hill-Kapturczak N, Truong L, Thamilselvan V, Visner GA, Nick HS, Agarwal A. (2000) Smad7-dependent regulation of heme oxygenase-1 by transforming growth factor-beta in human renal epithelial cells. *J Biol Chem.*; **275**: 40904–9.
- Hill-Kapturczak N, Chang SH, Agarwal A. (2002) Heme oxygenase and the kidney. *DNA Cell Biol.*; **21**: 307–21.
- Huang LE, Willmore WG, Gu J, Goldberg MA, Bunn HF. (1999) Inhibition of hypoxia-inducible factor 1 activation by carbon monoxide and nitric oxide. Implications

- for oxygen sensing and signaling. *J Biol Chem.*; **274**: 9038–44.
- Ingi T, Cheng J, Ronnett GV. (1996) Carbon monoxide: an endogenous modulator of the nitric oxide-cyclic GMP signaling system. *Neuron.*; **16**: 835–42.
- Ishikawa K, Maruyama Y. (2001) Heme oxygenase as an intrinsic defense system in vascular wall: implication against atherogenesis. *J Atheroscler Thromb.*; **8**: 63–70.
- Ishikawa K, Sugawara D, Goto J, Watanabe Y, Kawamura K, Shiomi M, Itabe H, Maruyama Y. (2001a) Heme oxygenase-1 inhibits atherogenesis in Watanabe heritable hyperlipidemic rabbits. *Circulation.*; **104**: 1831–6.
- Ishikawa K, Sugawara D, Wang XP, Suzuki K, Itabe H, Maruyama Y, Lusis AJ. (2001b) Heme oxygenase-1 inhibits atherosclerotic lesion formation in LDL-receptor knockout mice. *Circ Res.*; **88**: 506–12.
- Jeney V, Balla J, Yachie A, Varga Z, Vercellotti GM, Eaton JW, Balla G. (2002) Pro-oxidant and cytotoxic effects of circulating heme. *Blood.*; **100**: 879–87.
- Jozkowicz A, Huk I, Nigisch A, Weigel G, Dietrich W, Motterlini R, Dulak J. (2003) Heme oxygenase and angiogenic activity of endothelial cells: stimulation by carbon monoxide, inhibition by tin protoporphyrin-IX. *Antioxid Redox Signal*; in press.
- Juan SH, Lee TS, Tseng KW, Liou JY, Shyue SK, Wu KK, Chau LY. (2001) Adenovirus-mediated heme oxygenase-1 gene transfer inhibits the development of atherosclerosis in apolipoprotein E-deficient mice. *Circulation.*; **104**: 1519–25.
- Karlsson JO, Axelsson KL, Andersson RG. (1985) Effects of hydroxyl radical scavengers KCN and CO on ultraviolet light-induced activation of crude soluble guanylate cyclase. *J Cyclic Nucleotide Protein Phosphor Res.*; **10**: 309–15.
- Kawashima A, Oda Y, Yachie A, Koizumi S, Nakanishi I. (2002) Heme oxygenase-1 deficiency: the first autopsy case. *Hum Pathol.*; **33**: 125–30.
- Ke B, Shen XD, Melinek J, Gao F, Ritter T, Volk HD, Busuttil RW, Kupiec-Weglinski JW. (2001) Heme oxygenase-1 gene therapy: a novel immunomodulatory approach in liver allograft recipients? *Transplant Proc.*; **33**: 581–2.
- Ke B, Buelow R, Shen XD, Melinek J, Amersi F, Gao F, Ritter T, Volk HD, Busuttil RW, Kupiec-Weglinski JW. (2002) Heme oxygenase 1 gene transfer prevents CD95/Fas ligand-mediated apoptosis and improves liver allograft survival via carbon monoxide signaling pathway. *Hum Gene Ther.*; **13**: 1189–99.
- Ke B, Shen XD, Zhai Y, Gao F, Busuttil RW, Volk HD, Kupiec-Weglinski JW. (2002) Heme oxygenase 1 mediates the immunomodulatory and antiapoptotic effects of interleukin 13 gene therapy in vivo and in vitro. *Hum Gene Ther.*; **13**: 1845–57.
- Kharitonov VG, Sharma VS, Pilz RB, Magde D, Koesling D. (1995) Basis of guanylate cyclase activation by carbon monoxide. *Proc Natl Acad Sci U S A.*; **92**: 2568–71.
- Kharitonov SA, Barnes PJ. (2002) Biomarkers of some pulmonary diseases in exhaled breath. *Biomarkers.*; **7**: 1–32.
- Kibbe MR, Li J, Nie S, Watkins SC, Lizonova A, Kovesdi I, Simmons RL, Billiar TR, Tzeng E. (2000) Inducible nitric oxide synthase (iNOS) expression upregulates p21 and inhibits vascular smooth muscle cell proliferation through p42/44 mitogen-activated protein kinase activation and independent of p53 and cyclic guanosine monophosphate. *J Vasc Surg.*; **31**: 1214–28.
- Koehler RC, Traystman RJ. (2002) Cerebrovascular effects of carbon monoxide. *Antioxid Redox Signal.*; **4**: 279–90.
- Kourembanas S. (2002) Hypoxia and carbon monoxide in the vasculature. *Antioxid Redox Signal.*; **4**: 291–9.
- Kozma F, Johnson RA, Nasjletti A. (1997) Role of carbon monoxide in heme-induced vasodilation. *Eur J Pharmacol.*; **323**: R1–2.

- Kozma F, Johnson RA, Zhang F, Yu C, Tong X, Nasjletti A. (1999) Contribution of endogenous carbon monoxide to regulation of diameter in resistance vessels. *Am J Physiol.*; **276**: R1087-94.
- Kutty RK, Nagineni CN, Kutty G, Hooks JJ, Chader GJ, Wiggert B. (1994) Increased expression of heme oxygenase-1 in human retinal pigment epithelial cells by transforming growth factor-beta. *J Cell Physiol.*; **159**: 371-8.
- Lautier D, Luscher P, Tyrrell RM. (1992) Endogenous glutathione levels modulate both constitutive and UVA radiation/hydrogen peroxide inducible expression of the human heme oxygenase gene. *Carcinogenesis.*; **13**: 227-32.
- Lee TS, Chau LY. (2002) Heme oxygenase-1 mediates the anti-inflammatory effect of interleukin-10 in mice. *Nat Med.*; **8**: 240-6.
- Li Volti G, Wang J, Traganos F, Kappas A, Abraham NG. (2002) Differential effect of heme oxygenase-1 in endothelial and smooth muscle cell cycle progression. *Biochem Biophys Res Commun.*; **296**: 1077-82.
- Liu Y, Christou H, Morita T, Laughner E, Semenza GL, Kourembanas S. (1998) Carbon monoxide and nitric oxide suppress the hypoxic induction of vascular endothelial growth factor gene *via* the 5' enhancer. *J Biol Chem.*; **273**: 15257-62.
- Liu XM, Chapman GB, Wang H, Durante W. (2002) Adenovirus-mediated heme oxygenase-1 gene expression stimulates apoptosis in vascular smooth muscle cells. *Circulation.*; **105**: 79-84.
- Maines MD. (1997) The heme oxygenase system: a regulator of second messenger gases. *Annu Rev Pharmacol Toxicol.*; **37**: 517-54.
- Malaguarnera L, Pilastro MR, Quan S, Ghattas MH, Yang L, Mezentssev AV, Kushida T, Abraham NG, Kappas A. (2002) Significance of heme oxygenase in prolactin-mediated cell proliferation and angiogenesis in human endothelial cells. *Int J Mol Med.*; **10**: 433-40.
- Marshall HE, Merchant K, Stamler JS. (2000) Nitrosation and oxidation in the regulation of gene expression. *FASEB J.*; **14**: 1889-900.
- Maxwell PH, Ratcliffe PJ. (2002) Oxygen sensors and angiogenesis. *Semin Cell Dev Biol.*; **13**: 29-37.
- McCoubrey WK Jr, Huang TJ, Maines MD. (1997) Isolation and characterization of a cDNA from the rat brain that encodes hemoprotein heme oxygenase-3. *Eur J Biochem.*; **247**: 725-32.
- Meister A. (1994) Glutathione-ascorbic acid antioxidant system in animals. *J Biol Chem.*; **269**: 9397-400.
- Morita T, Kourembanas S. (1995) Endothelial cell expression of vasoconstrictors and growth factors is regulated by smooth muscle cell-derived carbon monoxide. *J Clin Invest.*; **96**: 2676-82.
- Morita T, Perrella MA, Lee ME, Kourembanas S. (1995) Smooth muscle cell-derived carbon monoxide is a regulator of vascular cGMP. *Proc Natl Acad Sci U S A.*; **92**: 1475-9.
- Morita T, Mitsialis SA, Koike H, Liu Y, Kourembanas S. (1997) Carbon monoxide controls the proliferation of hypoxic vascular smooth muscle cells. *J Biol Chem.*; **272**: 32804-9.
- Motterlini R, Clark JE, Foresti R, Sarathchandra P, Mann BE, Green CJ. (2002) Carbon monoxide-releasing molecules: characterization of biochemical and vascular activities. *Circ Res.*; **90**: E17-24.
- Ohta K, Yachie A, Fujimoto K, Kaneda H, Wada T, Toma T, Seno A, Kasahara Y, Yokoyama H, Seki H, Koizumi S. (2000) Tubular injury as a cardinal pathologic feature in human heme oxygenase-1 deficiency. *Am J Kidney Dis.*; **35**: 863-70.
- Otterbein LE, Choi AM. (2000) Heme oxygenase: colors of defense against cellular stress. *Am J Physiol Lung Cell Mol Physiol.*; **279**: L1029-37.
- Otterbein LE, Bach FH, Alam J, Soares M, Tao Lu H, Wysk M, Davis RJ, Flavell RA, Choi AM. (2000) Carbon monoxide has anti-inflammatory effects involving the mitogen-activated protein kinase pathway. *Nat Med.*; **6**: 422-8.

- Peyton KJ, Reyna SV, Chapman GB, Ensenat D, Liu XM, Wang H, Schafer AI, Durante W. (2002) Heme oxygenase-1-derived carbon monoxide is an autocrine inhibitor of vascular smooth muscle cell growth. *Blood.*; **99**: 4443–8.
- Piantadosi CA. (2002) Biological chemistry of carbon monoxide. *Antioxid Redox Signal.*; **4**: 259–70.
- Poss KD, Tonegawa S. (1997a) Heme oxygenase 1 is required for mammalian iron reutilization. *Proc Natl Acad Sci U S A.*; **94**: 10919–24.
- Poss KD, Tonegawa S. (1997b) Reduced stress defense in heme oxygenase 1-deficient cells. *Proc Natl Acad Sci U S A.*; **94**: 10925–30.
- Quan S, Yang L, Shenouda S, Jiang H, Balazy M, Schwartzman ML, Shibahara I, Shinohara K, Abraham NG. (2002) Functional expression of human heme oxygenase-1 (HO-1) driven by HO-1 promoter *in vitro* and *in vivo*. *J Cell Biochem.*; **85**: 410–21.
- Rotenberg MO, Maines MD. (1990) Isolation, characterization, and expression in *Escherichia coli* of a cDNA encoding rat heme oxygenase-2. *J Biol Chem.*; **265**: 7501–6.
- Rutter J, Reick M, McKnight SL. (2002) Metabolism and the control of circadian rhythms. *Annu Rev Biochem.*; **71**: 307–31.
- Sato K, Balla J, Otterbein L, Smith RN, Brouard S, Lin Y, Csizmadia E, Seigny J, Robson SC, Vercellotti G, Choi AM, Bach FH, Soares MP. (2001) Carbon monoxide generated by heme oxygenase-1 suppresses the rejection of mouse-to-rat cardiac transplants. *J Immunol.*; **166**: 4185–94.
- Saunders EL, Maines MD, Meredith MJ, Freeman ML. (1991) Enhancement of heme oxygenase-1 synthesis by glutathione depletion in Chinese hamster ovary cells. *Arch Biochem Biophys.*; **288**: 368–73.
- Scapagnini G, D'Agata V, Colombrita C, Caruso A, Quattrone A, Calabrese V, Cavallaro S. (2002) Regional distribution of HO-3 in rat brain: peculiarities of a probable retrotransposed gene. *2nd International Conference on Heme Oxygenase (HO/CO)*, Catania, Sicily, June 2002, abstract 155.
- Semenza G. (2002) Signal transduction to hypoxia-inducible factor 1. *Biochem Pharmacol.*; **64**: 993–8.
- Shi W, Haberland ME, Jien ML, Shih DM, Lusis AJ. (2000a) Endothelial responses to oxidized lipoproteins determine genetic susceptibility to atherosclerosis in mice. *Circulation.*; **102**: 75–81.
- Shi W, Wang NJ, Shih DM, Sun VZ, Wang X, Lusis AJ. (2000b) Determinants of atherosclerosis susceptibility in the C3H and C57BL/6 mouse model: evidence for involvement of endothelial cells but not blood cells or cholesterol metabolism. *Circ Res.*; **86**: 1078–84.
- Shibahara S, Muller R, Taguchi H, Yoshida T. (1985) Cloning and expression of cDNA for rat heme oxygenase. *Proc Natl Acad Sci U S A.*; **82**: 7865–9.
- Shih DM, Xia YR, Wang XP, Miller E, Castellani LW, Subbanagounder G, Cheroutre H, Faull KF, Berliner JA, Witztum JL, Lusis AJ. (2000) Combined serum paraoxonase knockout/apolipoprotein E knockout mice exhibit increased lipoprotein oxidation and atherosclerosis. *J Biol Chem.*; **275**: 17527–35.
- Sjöstrand T. (1952) The formation of carbon monoxide by the decomposition of haemoglobin *in vivo*. *Acta Physiol Scand.*; **26**: 338.
- Snyder SH, Jaffrey SR, Zakhary R. (1998) Nitric oxide and carbon monoxide: parallel roles as neural messengers. *Brain Res Brain Res Rev.*; **26**: 167–75.
- Snyder SH, Baranano DE. (2001) Heme oxygenase: a font of multiple messengers. *Neuropsychopharmacology.*; **25**: 294–8.
- Soares MP, Lin Y, Anrather J, Csizmadia E, Takigami K, Sato K, Grey ST, Colvin RB, Choi AM, Poss KD, Bach FH. (1998) Expression of heme oxygenase-1 can determine cardiac xenograft survival. *Nat Med.*; **4**: 1073–7.
- Stocker R, Glazer AN, Ames BN. (1987a) Antioxidant activity of albumin-bound bilirubin. *Proc Natl Acad Sci U S A.*; **84**: 5918–22.

- Stocker R, Yamamoto Y, McDonagh AF, Glazer AN, Ames BN. (1987b) Bilirubin is an antioxidant of possible physiological importance. *Science*; **235**: 1043–6.
- Stupfel M, Bouley G. (1970) Physiological and biochemical effects on rats and mice exposed to small concentrations of carbon monoxide for long periods. *Ann NY Acad Sci*; **174**: 342–68.
- Suematsu M, Goda N, Sano T, Kashiwagi S, Egawa T, Shinoda Y, Ishimura Y. (1995) Carbon monoxide: an endogenous modulator of sinusoidal tone in the perfused rat liver. *J Clin Invest*; **96**: 2431–7.
- Tenhunen R, Marver HS, Schmid R. (1968) The enzymatic conversion of heme to bilirubin by microsomal heme oxygenase. *Proc Natl Acad Sci U S A*; **61**: 748–55.
- Tenhunen R, Marver HS, Schmid R. (1969) Microsomal heme oxygenase. Characterization of the enzyme. *J Biol Chem*; **244**: 6388–94.
- Tenhunen R, Marver HS, Schmid R. (1970) The enzymatic catabolism of hemoglobin: stimulation of microsomal heme oxygenase by hemin. *J Lab Clin Med*; **75**: 410–21.
- Thiemermann C. (2001) Inhaled CO: deadly gas or novel therapeutic? *Nat Med*; **7**: 534–5.
- Thom SR, Fisher D, Xu YA, Garner S, Ischiropoulos H. (1999) Role of nitric oxide-derived oxidants in vascular injury from carbon monoxide in the rat. *Am J Physiol*; **276**: H984–92.
- Thom SR, Fisher D, Xu YA, Notarfrancesco K, Ischiropoulos H. (2000) Adaptive responses and apoptosis in endothelial cells exposed to carbon monoxide. *Proc Natl Acad Sci U S A*; **97**: 1305–10.
- Thorup C, Jones CL, Gross SS, Moore LC, Goligorsky MS. (1999) Carbon monoxide induces vasodilation and nitric oxide release but suppresses endothelial NOS. *Am J Physiol*; **277**: F882–9.
- Togane Y, Morita T, Suematsu M, Ishimura Y, Yamazaki JI, Katayama S. (2000) Protective roles of endogenous carbon monoxide in neointimal development elicited by arterial injury. *Am J Physiol Heart Circ Physiol*; **278**: H623–32.
- Tzeng E, Kim YM, Pitt BR, Lizonova A, Kovesdi I, Billiar TR. (1997) Adenoviral transfer of the inducible nitric oxide synthase gene blocks endothelial cell apoptosis. *Surgery*; **122**: 255–63.
- Utz J, Ullrich V. (1991) Carbon monoxide relaxes ileal smooth muscle through activation of guanylate cyclase. *Biochem Pharmacol*; **41**: 1195–201.
- Vercellotti GM, Balla G, Balla J, Nath K, Eaton JW, Jacob HS. (1994) Heme and the vasculature: an oxidative hazard that induces antioxidant defenses in the endothelium. *Artif Cells Blood Substit Immobil Biotechnol*; **22**: 207–13.
- Vreman HJ, Wong RJ, Stevenson DK. (2002) Sources, sinks and measurement of carbon monoxide. In: *Carbon monoxide and cardiovascular functions*. Wang R, ed, pp 45–65. CRC Press, Boca Raton, London, New York, Washington.
- Wang R. (1998) Resurgence of carbon monoxide: an endogenous gaseous vasorelaxing factor. *Can J Physiol Pharmacol*; **76**: 1–15.
- Weber CM, Eke BC, Maines MD. (1994) Corticosterone regulates heme oxygenase-2 and NO synthase transcription and protein expression in rat brain. *J Neurochem*; **63**: 953–62.
- Yachie A, Niida Y, Wada T, Igarashi N, Kaneda H, Toma T, Ohta K, Kasahara Y, Koizumi S. (1999) Oxidative stress causes enhanced endothelial cell injury in human heme oxygenase-1 deficiency. *J Clin Invest*; **103**: 129–35.
- Yoshida T, Biro P, Cohen T, Muller RM, Shibahara S. (1988) Human heme oxygenase cDNA and induction of its mRNA by hemin. *Eur J Biochem*; **171**: 457–61.
- Zhang J, Piantadosi CA. (1992) Mitochondrial oxidative stress after carbon monoxide hypoxia in the rat brain. *J Clin Invest*; **90**: 1193–9.
- Zhang F, Kaide JI, Rodriguez-Mulero F, Abraham NG, Nasjletti A. (2001) Vasoregulatory

function of the heme-heme oxygenase-carbon monoxide system. *Am J Hypertens.*; **14**: 62S–7S.

Zhang M, Zhang BH, Chen L, An W. (2002) Overexpression of heme oxygenase-1 protects smooth muscle cells against oxidative injury and inhibits cell proliferation. *Cell Res.*; **12**: 123–32.